

### **Faxitron X-ray examine the skeleton:**

The faxitron is located in room 512.

Get Key from Tina Kilts (room 227)

Turn on machine to warm up (takes 2-5 min).

Typical exposure for mouse bones: Time 90 second (enter), KV 30 (enter)  
(may need changes depending on what you are imaging)

Prepare mouse by treatment with 0.1cc adwertin/10gm body weigh or by  
CO2 treatment. Use tape in place mouse onto a plastic surface

Use Kodak XTL2 film (type that says for therapy localization)

Sample closest to the top will be 5x, the next level is 4X etc. Samples on  
lowest level are 1X

Expose sample

Develop film to determine if exposure is correct

Often several views are taken (dorsal ventral, ventral dorsal, isolated heads  
or legs etc)

### **Histology Preparation:**

Dissect tissues from the mouse and incubate in 4% PFA in PBS for 16 hours  
followed with storage at 4 C in PBS or 70% ethanol.

10% formaldehyde is used for head samples (two weeks) and is located in  
512. Commercially available Zeta Fix can also used for sample fixation (this  
fixative is stable and does not need to be made fresh like the 4%PFA). For  
plastic: do not decalcify. For normal H and E: decalcify for two- four weeks  
in immunocal changing solutions 2x./week. Test decalcification by  
cutting/Faxitroning. Post-fix in Z-fix 3 days, wash in water, store 4 C in 70%  
ethanol

Send samples Histoserve: 301-963-2606 (contact person: Pratiba) with clear  
graphic instructions.

Bood analysis: Analytics, 301-921-0168